

Bioremediation Potentials of Heavy Metal Tolerant Bacteria Isolated from Petroleum Refinery Effluent

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Abstract: Five heavy metals tolerant bacteria were isolated from petroleum refinery effluent and identified as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *E. coli*, *Proteus vulgaris* and *Klebsiella pneumoniae*. Each isolate was inoculated into different concentrations of cadmium, chromium, nickel and zinc to determine its maximum tolerance for each heavy metal. All five isolates had low maximum tolerance concentration for cadmium (0.9 mg/L) when compared to chromium (5 mg/L), nickel (5 mg/L) and zinc (7 mg/L). Mixed culture consortium (MCC) remediated high percentage of cadmium (100%), chromium (33.4%), nickel (73.9%), and zinc (90.1%) from the petroleum refinery effluent than pure culture isolates. Among the pure culture isolates *Pseudomonas aeruginosa* (Cd (100%), Cr (23.1%), Ni (64.3%) & Zn (53.9%)) yielded high values for the reduction of heavy metals in the refinery effluent when compared to *Staphylococcus aureus*, *E.coli*, *Proteus vulgaris* and *Klebsiella pneumoniae*. The isolated bacteria were effective for the remediation of heavy metals from petroleum refinery effluent.

Keywords: Bioremediation, Heavy Metal, Bacteria, Refinery, Petroleum

1. Introduction

Industrial development results in the generation of industrial effluents and if untreated, results in water, sediments and soil pollution (Fakayode, 2005). Industrial effluent is heavily loaded with different types of organic and inorganic pollutants, which are discharged into receiving water bodies (Jern, 2006).

Liquid wastes generated by refineries do compose of chemical components such as oil and grease, phenols, Benzene, Toluene, Ethyl benzene and Xylene (BTEX), ammonia, suspended solids, cyanide, sulfide, nitrogen compounds and heavy metals such as Iron (Fe), Cadmium (Cd), Nickel (Ni), Chromium (Cr), Copper (Cu), Molybdenum (Mo), Selenium (S), Vanadium (V) and Zinc (Zn) (Jorge *et al.*, 2011).

Heavy metal is a general collective term that applies to the group of metals and metalloids with atomic density greater

than 4000 kgm⁻³, or 5 times more than water (Garbarino *et al.*, 1995). Some heavy metals are purely toxic with no known role (Shi *et al.*, 2002) while others are essential for life at low concentration but become toxic at high concentrations (Franke *et al.*, 2003).

Conventional physicochemical techniques for metal remediation such as filtration, acid leaching, electrochemical processes or ion exchange are expensive and may not be very effective. Biological treatment is preferred over physicochemical for toxic pollutants as the former is cost effective, efficient and environmentally friendlier (Ojo 2006; Hamza *et al.*, 2009). Recently, studies have discovered the use of microorganisms, plants and enzymes extracted from other organisms for the remediation of pollutants in contaminated soil, water and ground water (Hamza *et al.*, 2009; Usman *et al.*, 2012).

Mechanisms for metal removal by bacteria include accumulation, precipitation, complexation, volatilization, adsorption and enzyme degradation (Dubey, 2012). *Bacillus spp*, *Pseudomonas aeruginosa*, *Citrobacters pp*, *Zoogleas pp*, *Chlorella vulgaris* have been identified as microbes that can utilize heavy metals (Sar *et al.*, 1999; Philip *et al.*, 2000; Sar and D'Souza, 2001; Gunasekaran *et al.*, 2003). The ability of microbial strains to develop in the presence of heavy metals would be helpful in wastewater treatment where microorganisms are directly involved in bioremediation of the wastewater (Munoz *et al.*, 2006).

2. Materials and Methods

2.1. Sample Collection

Effluent samples were collected from five different discharge points of the Kaduna Refining and Petrochemical Company (KRPC) into sterilized bottles and syringes. The samples were transported to the Department of Biological Sciences Laboratory, Nigerian Defence Academy (NDA), Kaduna for bacteria isolation.

2.2. Isolation of Possible Heavy Metals Tolerant Bacteria

To 0.1ml of the 10-fold diluted effluent samples were each inoculated using pour plate method into nutrient agar plates containing a combined concentrations of the heavy metals in their salt which include potassium dichromate ($K_2Cr_2O_7$), cadmium nitrate $Cd(NO_3)_2$, nickel chloride ($NiCl_2$) and zinc sulphate ($ZnSO_4$). For each nutrient plate, a concentration of 0.5 mg/ml was incorporated. The plates were incubated at room temperature for 24-48 hours. The bacteria were isolated and subcultured into nutrient plates to obtain pure cultures of each possible tolerant strain (Pandit *et al.*, 2013).

2.3. Test for Tolerance Trend of Bacteria Isolates to Various Concentrations of each Heavy Metal

Each isolated bacteria was inoculated into a test tube containing 10ml nutrient broth at room temperature for 24 hours. Twenty microliters (20 μ l) of the isolated bacteria were each inoculated into a nutrient broth containing different concentrations of heavy metals; 0.5 mg/ml, 1 mg/ml, 2 mg/ml, 4 mg/ml, 6 mg/ml and 8 mg/ml respectively for Cr, Ni and Zn while the concentrations for Cadmium were 0.5 mg/ml, 0.6 mg/ml, 0.7 mg/ml, 0.8 mg/ml, 0.9 mg/ml and 1 mg/ml. The inhibitory concentrations were measured using a spectrophotometer at an absorbance of 600 nm against a nutrient broth (blank) containing the same amount of heavy metals (Pandit *et al.*, 2013).

2.4. Identification of Isolated Bacteria

Isolated colonies of the heavy metal tolerant bacteria were observed for identification based on their superficial forms (circular, filamentous and irregular); elevation (Flat, convex and umbonate), margin and shape (spiral, rod or cocci). Gram staining (Aneja, 2006) and reactions to biochemical

tests were used. The results obtained were compared with bergey's manual of systematic bacteriology for the identification (Holt *et al.*, 1994).

2.5. Evaluation of the Potentials of Heavy Metals by Bacterial Isolates in Effluent

The wastewater was sterilized using an autoclave at 121 °C for 15mins. To seven 120ml sterile conical flasks was added 100 ml of the sterilized wastewater each. The first flask was not inoculated with bacterium (Control), the second, third, fourth, fifth and sixth were each inoculated with 20 μ l of a 24hrs nutrient broth culture of *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *E.coli*, *Proteus vulgaris* and *Klebsiella pneumoniae* respectively. The seventh flask was inoculated with a mixed culture consortium (MCC) of all the five bacteria. The experiment was left to stand for 24hrs after which the concentration of cadmium, chromium, nickel and zinc was measured using atomic absorption spectrophotometer. Percentage reductions of heavy metals were observed when comparison was done between the values before and after treatment.

3. Results

Gram staining of each colony of the heavy metal tolerant bacteria revealed isolates A, C, D, and E as gram negative bacteria and Isolates B gram positive bacteria. Biochemical characterization (Table 1) of Isolates A, B, C, D and E revealed them as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *E.coli*, *Proteus vulgaris* and *Klebsiella pneumoniae*. The isolates showed different ranges of heavy metals tolerance using various concentration of each metal (Figure 1, 2, 3 and 4). The microbial turbidity decreased with increase in concentration of heavy metals (0.5- 8 mg/ml). The Optical Density (OD) values recorded for *Pseudomonas aeruginosa* had Maximum Tolerance Concentration (MTC) of 0.9 mg/L for cadmium, 4 mg/L for chromium, 4 mg/L for nickel and 7 mg/L for zinc. *Staphylococcus aureus* had MTC of 0.8 mg/L for cadmium, 3 mg/L for chromium, 3 mg/L for nickel and 4 mg/L for zinc. *Escherichia coli* had MTC of 0.8mg/L for cadmium, 5 mg/L for chromium, 3 mg/L for nickel and 6 mg/L for zinc. *Proteus vulgaris* had MTC of 0.6 mg/L for cadmium, 5 mg/L for chromium, 5 mg/L for nickel and 6 mg/L for zinc. *Klebsiella pneumoniae* had MTC of 0.7 mg/L for cadmium, 4 mg/L for chromium, 2 mg/L for nickel and 5 mg/L for zinc. For cadmium (Fig. 1) *Pseudomonas aeruginosa* had maximum tolerance while *Proteus vulgaris* had the minimum tolerance. Chromium (Fig. 2) was highly tolerable by *E.coli* and *Proteus vulgaris* and least by *Staphylococcus aureus*. Nickel (Fig. 3) had maximum tolerance by *Proteus vulgaris* and *Pseudomonas aeruginosa* and minimum tolerance by *Klebsiella pneumoniae*. Zinc (Fig. 4) was highly tolerable by *Pseudomonas aeruginosa* and least by *Staphylococcus aureus*.

Zinc concentration was reduced by *Pseudomonas aeruginosa* 53.9%, *Staphylococcus aureus* 46.6%, *E.coli*

88%, *Proteus vulgaris* 89%, *Klebsiella pneumonia* 20%, and Mixed culture Consortium (MCC) 90.1%. Chromium concentration was reduced by *Pseudomonas aeruginosa* 23.1%, *Staphylococcus aureus* 11.6%, *E.coli* 21.6%, *Proteus vulgaris* 20%, *Klebsiella pneumonia* 15.4%, and Mixed Culture Consortium (MCC) 33.14%. Nickel concentration was reduced by *Pseudomonas aeruginosa* 64.3%,

Staphylococcus aureus 44.7%, *E.coli* 54.4%, *Proteus vulgaris* 60.47%, *Klebsiella pneumonia* 56.3%, and Mixed Culture Consortium (MCC) 73.91%. Cadmium concentration was reduced by *Pseudomonas aeruginosa* 100%, *Staphylococcus aureus* 36.1%, *E.coli* 13.1%, *Proteus vulgaris* 65%, *Klebsiella pneumonia* 69.6% and Mixed Culture Consortium (MCC) 100%.

Table 1. Biochemical and Morphological Characterization of the Heavy Metal Tolerant Bacteria Isolates from Petroleum Refinery Effluent.

Colonial Variables	Cultural and Colony Morphologies				
	IST A	IST B	IST C	IST D	IST E
Shape	Rod	Cocci	Rod	Rod	Rod
Colour	Green pigment	White	White	Yellow	Gray
Form	Circular	Smooth	Circular	Circular	Irregular
Elevation	Convex	Flat	Convex	Raised	Raised
Gram staining	-	+	-	-	-
Biochemical Reactivities					
Indole	-	-	+	+	-
Methyl red	-	+	+	+	-
Voges Proskauer	+	-	-	+	+
Citrate	+	+	-	-	+
Urease	-	-	-	+	+
Catalase	+	+	+	+	+
Oxidase	+	+	-	-	-
Ornithine decarboxylase	-	-	-	-	-
Mannitol	+	+	+	-	+
Nitrate reduction	+	+	-	+	+
H ₂ S production	-	-	-	+	-
Glucose	-	-	+	+	+
Lactose	-	+	+	-	+
Sucrose	-	-	-	+	-
Gas production	-	-	+	+	+
Motility	+	-	+	+	+
Probable Bacteria Isolates	<i>P. aeruginosa</i>	<i>S. aureus</i>	<i>E. coli</i>	<i>P. vulgaris</i>	<i>K. Pneumoniae</i>

Key: IST- Bacteria Isolates, + Positive, - Negative, *P. aeruginosa* - *Pseudomonas aeruginosa*, *S. aureus*- *Staphylococcus aureus*, *E. coli* -*Escherichia coli*, *P. vulgaris* - *Proteus vulgaris* and *K. Pneumoniae* – *Klebsiella pneumoniae*.

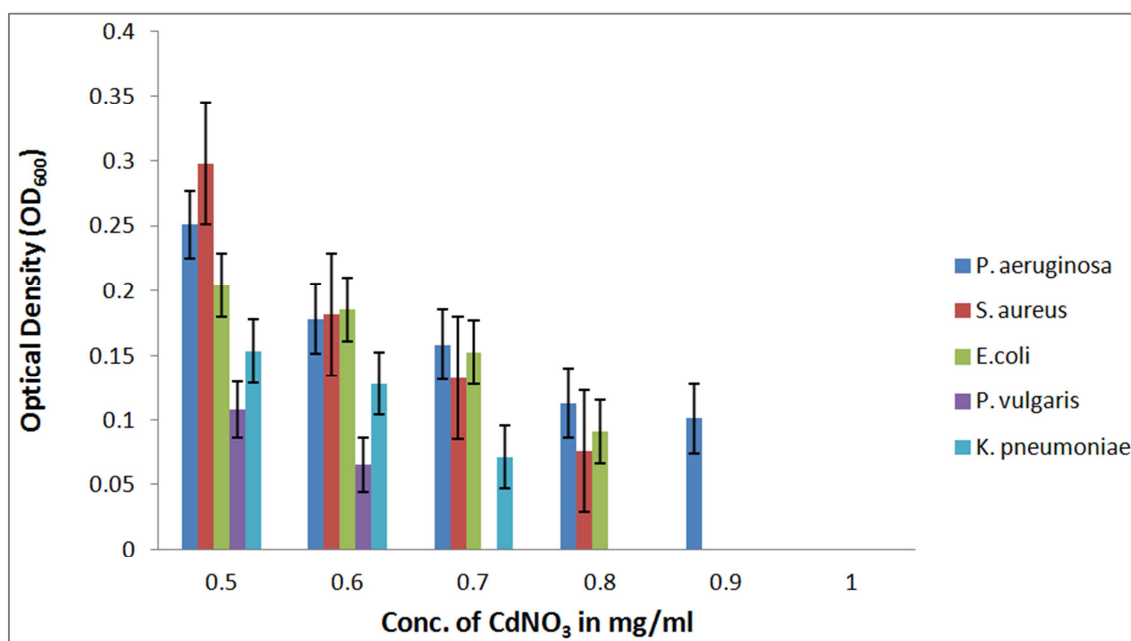


Fig. 1. Tolerance Trend of Bacteria Isolates from Refinery Effluent to Variable concentrations of CdNO₃.

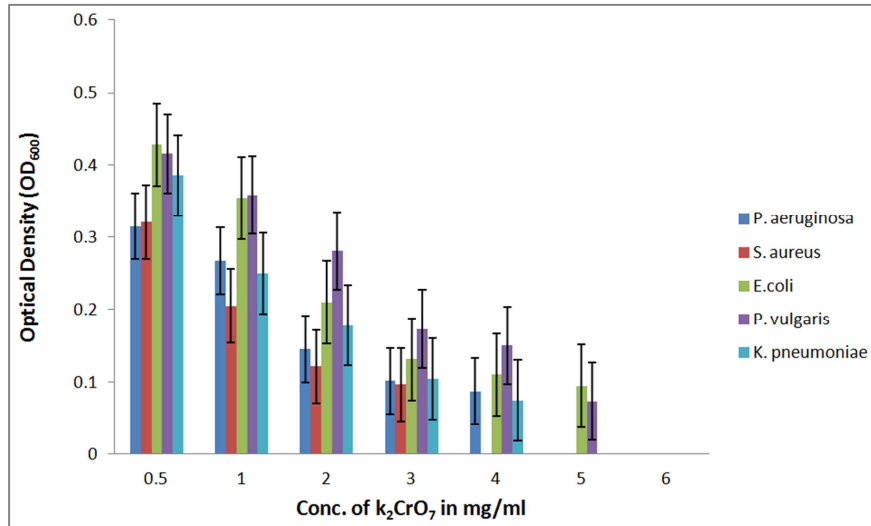


Fig. 2. Tolerance Trend of Bacteria Isolates from Refinery Effluent to Variable concentrations of K₂CrO₇.

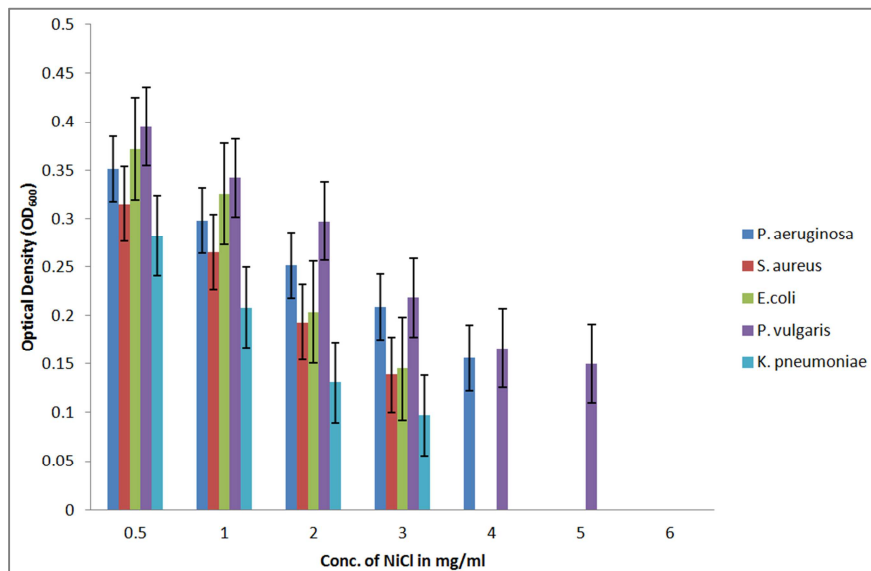


Fig. 3. Tolerance Trend of Bacteria Isolates from Refinery Effluent to Variable Concentrations of NiCl₂.

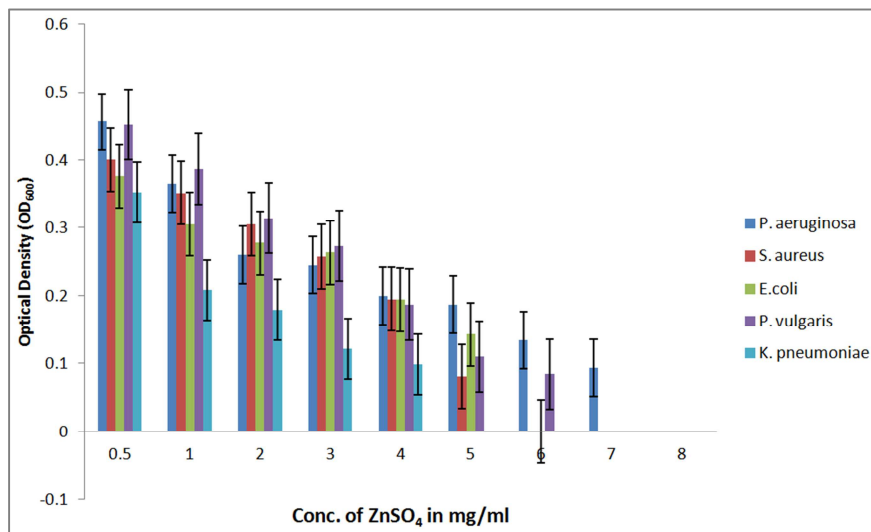


Fig. 4. Tolerance Trend of Bacteria Isolates from Refinery Effluent to Variable Concentrations of ZnSO₄.

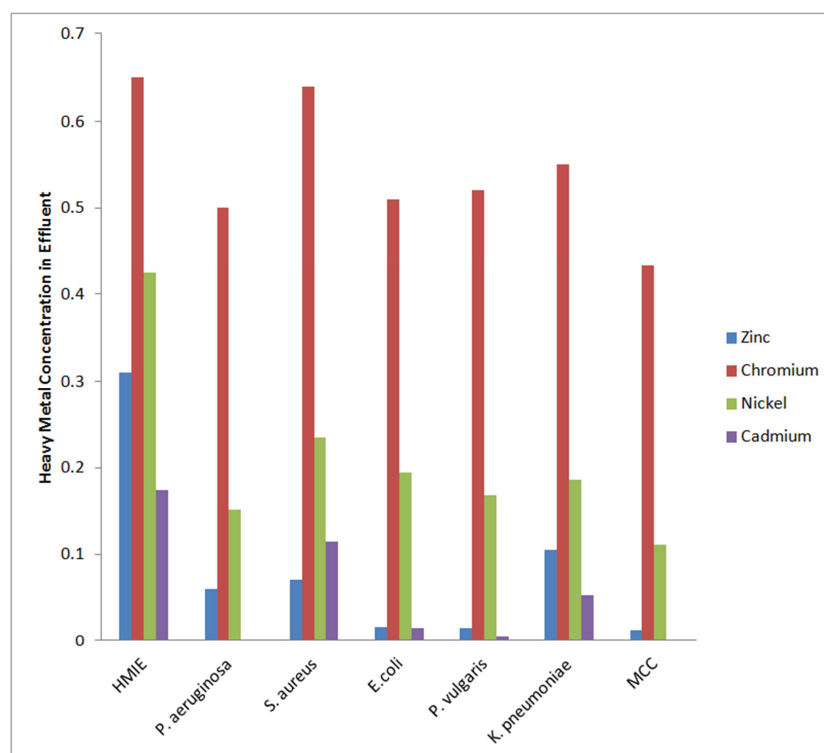


Fig. 5. Heavy Metals reduction by bacteria isolates in KRPC effluent; MCC – Mixed Culture Consortium (Consisting of all five isolates), HMIE- Heavy Metal in Effluent.

4. Discussion

Five bacterial isolates with maximum tolerance trend for cadmium, chromium, nickel and zinc were identified as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Escherichia coli*, *Proteus vulgaris* and *Klebsiella pneumoniae* (Table 1). This work conformed to Rajibanshi (2008) who isolated *Staphylococcus* spp., *E. coli*, *Klebsiella* spp., *Actinobacter* spp., *Flavobacterium* spp., *Citrobacter* spp., *Pseudomonas* spp., *Methylbacterium* spp. as heavy metal tolerant bacteria from a sewage treatment plant in Guheswori, Nepali. Similarly, Mgbema *et al.* (2012) isolated *Bacillus* spp., *E. coli*, *Micrococcus* spp., *Proteus* spp., *Pseudomonas* spp. and *Aeromonas* spp. as heavy metal resistance bacteria from Otamiri River, Owerri. Among the five bacteria isolates, *Pseudomonas aeruginosa* showed maximum tolerance to heavy metals. This finding is in agreement with Leedjaru *et al.* (1996) who reported that *Pseudomonas aeruginosa* contains at least four Zinc or Cadmium or Lead efflux transporters and two Cadmium, Zinc and Cobalt (CZC) chemiosmotic transporters. *Proteus vulgaris* showed maximum tolerance to Nickel and Zinc. *Proteus* spp. had also been identified with high maximum tolerance to Zinc (2.25 mg/L) according to Mgbema *et al.* (2012).

The microbial turbidity decreased with increase in concentration of heavy metals (0.5-8 mg/ml) indicating toxic and inhibitory effect of the heavy metals on the growth of bacteria (Fig. 1, 2, 3 and 4). According to Konopka *et al.* (1999), the resistance mechanisms employed by the bacteria do not offer protection at extremely high levels of free metal

ions and a lethal toxic effect is observed above the critical level. Metal tolerance by the five heavy metals tolerant bacteria in decreasing order was Zn > Ni > Cr > Cd. The higher tolerance levels for Zinc and Nickel could be attributed to their classification as micronutrients that are needed by the bacteria in trace amount (Nies, 1999). According to European Union (2004), Nickel is used in some food supplements which contain micro-grams of Nickel per tablet. Cadmium's low tolerance by the five bacteria suggests the high toxic effect of this heavy metal which has been reported to be carcinogenic and could also lead to kidney dysfunction (WHO, 2003). Low tolerance to cadmium by the five bacteria isolates in this work among other metals is similar to the report made by Smrithi and Usha (2012) who reported decreasing order of metal tolerance in their research in the sequence Ni > Cr > Cu > Zn > and Cd for isolated heavy metal tolerant bacteria from tannery effluent in Tamil Nadu, India. But at variance from the report made by Amalesh *et al.* (2012) who stated that the decreasing order for metal tolerance is Cd > Cr > Ni > Co.

The heavy metals reduction by each bacteria isolates in the refinery wastewater revealed *P. aeruginosa* as the best heavy metal-tolerant isolate followed by *P. vulgaris*, *E. coli*, *K. pneumoniae* and *S. aureus* (Fig. 5). *Pseudomonas aeruginosa* showed 100% removal of Cadmium from the refinery wastewater. Momba and Ikonga (2013), in their research observed that *Pseudomonas putida* showed removal rates of 100% for Thallium, 96% for Lead, 83% for Vanadium, 71% for Cobalt, 57% for Nickel, 49% for Copper and 45% for Manganese. According to Vitri and Giownnetti (2008), *Pseudomonas* spp. has the ability to degrade heavy metals

present in industrial effluent. In general, the Mixed Culture Consortium (MCC) of the five heavy metal-tolerant bacteria showed the highest efficacy for heavy metal reduction in the effluents. The efficacy of removal of the four heavy metals in decreasing order was MCC>*P. aeruginosa*>*P. vulgaris*>*E.coli*>*K. pneumoniae*>*S. aureus*. The high bioremediation rate observed in MCC might be attributed to the synergy of metabolic activities by the consortium of five bacteria isolates due to the diversification of enzymatic types amongst the different types which might have enhanced the process than individual isolates with fewer enzymes types.

5. Conclusion

The five different bacteria isolates showed various level of maximum tolerance to different concentrations of cadmium, chromium, nickel and zinc. The efficacy of removal of the four heavy metals in decreasing order was MCC>*P. aeruginosa*>*P. vulgaris*>*E.coli*>*K. pneumoniae*>*S. aureus*. Overall, the mixed culture consortia had the highest percentages reduction of each heavy metal analyzed in the study. Hence, MCC are more effective for bioremediation of heavy metals than single colony bacterium.

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